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ance Notes on Codes and Abbreviations" appearing at the begin-
ning of each regular issue of the PCT Gazette.

(54) Title: SURFACTANT SYSTEM

(57) Abstract: The invention relates to an aqueous composition comprising in combination an alkyl pyrrolidone, such as a C8-C18 linear alkyl pyrrolidone, and an alkyl polysaccharide such as an alkyl polyglucoside, and a method of enhancing the efficacy of an enzyme containing composition for use in cleaning medical instruments comprising the step of including in said composition an alkyl pyrrolidone and an alkyl polysaccharide. The methods and compositions of the present invention may further include at least one enzyme such as proteases, lipases, amylases, and cellulases.

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SURFACTANT SYSTEM

TECHNICAL FIELD

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The invention relates to a surfactant system for use in aqueous compositions and more particularly to a surfactant system for use in, or addition to, aqueous enzyme based cleaning preparations. The invention has particular advantage when employed in enzyme based cleaning preparations intended for use for cleaning medical instruments.

BACKGROUND ART

20

Any discussion of the prior art throughout the specification should in no way be considered as an admission that such prior art is widely known or forms part of common general knowledge in the field.

25

The invention is herein described with particular reference to compositions for cleaning medical instruments, but is not limited to that use. As herein used the term "medical instruments" is intended broadly to include surgical instruments such as scalpels, biopsy instruments, clamps, and the like; endoscopes, colonoscopes, laparoscopes and other paraphernalia used for surgical investigation or intervention; and other instruments used in the practice of medicine, surgery, and dentistry which require cleaning, disinfection or sterilisation. The term is also intended to include instruments having similar cleaning requirements such as those used in hairdressing, cosmetic treatments, tattooing, body piercing etc as well as other applications where removal of human/animal sebum and/or body secretion is required. Many medical instruments are

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made from or incorporate a variety of construction materials and cleaning compositions must be such as not to attack any such materials.

It is common practice to clean medical instruments after use with one patient and prior to treatment of another by scrubbing to remove blood, tissue, loose proteinaceous and other soils, and then soaking for a predetermined period in an enzyme preparation adapted to further digest or loosen any proteinaceous materials remaining on the instrument surface. The instruments are then rinsed clean and subjected to further disinfection or sterilization procedures. To minimize the possibility of cross infection instruments used with one patient are desirably cleaned separately from those used with another.

Preparations for use in cleaning medical instruments consist generally of a concentrate comprising one or more surfactants in combination with one or more proteolytic enzymes. The concentrate is diluted to a working strength (normally a hundredfold dilution) prior to use. One such formulation has been described by NOVO NORDISK ("Novo-formulation"). Three preparations used internationally for this purpose are EPIZYME® a product of 3M, MEDIZYME® (a product of Whiteley Industries Pty Ltd., Sydney Australia) and ENDOZYME® (a product of Ruholf Corporation, NY, USA). While such products are generally effective and widely used there remains a need for products which are more efficient, and especially for preparations which are capable of achieving a predetermined level of effectiveness within a shorter time than those currently available.

The concentrate must be sufficiently stable in transit and storage prior to use to satisfy the needs of commerce. The choice of surfactants which in practice can be formulated in concentrated aqueous solutions is limited. Either the surfactants limit the storage stability of the concentrate, resulting in problems of foaming on dilution, provide insufficient reduction in surface tension, destabilize other components of the system such as enzymes, or suffer from other disadvantages. Mostly, formulators utilize surfactants chosen from a small group of conventional and well known nonionic surfactants which have been found to be satisfactory for such uses, but there remains an unsatisfied demand for formulations of improved efficiency.

It is an object of the invention to provide an improved surfactant system, and of preferred embodiments of the invention to provide an enzyme based cleaning

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composition which avoids problems of the prior art and/or provides improved performance.

DESCRIPTION OF THE INVENTION

According to a first aspect the invention provides an aqueous composition
5 comprising in combination an alkyl pyrrolidone and an alkyl polysaccharide.

For preference the alkyl pyrrolidone is a C8 - C18 linear alkyl pyrrolidone. N-dodecyl pyrrolidone ("N-DP") is highly preferred. Desirably, the alkyl polysaccharide is an alkyl polyglucoside ("APG").

According to a second aspect the invention provides a method of enhancing the
10 efficacy of an enzyme containing composition for use in cleaning medical instruments comprising the step of including an alkyl pyrrolidone and an alkyl polysaccharide.

Preferably the composition is a concentrate which contains surfactant according to the first aspect as part of its formulation, but the surfactant system can be added subsequently either to the concentrate or the working solution.

15 According to a third aspect the invention provides a method of cleaning a medical instrument including the step of treating the instrument with a composition including at least one enzyme, an alkyl pyrrolidone and an alkyl polysaccharide.

Unless the context clearly requires otherwise, throughout the description and the claims, the words 'comprise', 'comprising', and the like are to be construed in an
20 inclusive sense as opposed to an exclusive or exhaustive sense; that is to say, in the sense of "including, but not limited to".

Alkylpyrrolidones are known to reduce surface tension progressively with increasing concentration in water achieving a minimum surface tension (maximum effectiveness) at a concentration of about 0.1% in the case of dodecyl pyrrolidone and
25 about 0.002% in the case of octyl pyrrolidone, which concentrations correspond to the maximum solubility of the respective pyrrolidones in water. This has in the past inhibited their use in concentrates, since the maximum concentration in an aqueous system is inevitably less than fully effective as a surfactant when diluted to any substantial degree.

30 BEST MODES FOR CARRYING OUT THE INVENTION

An additive according to the invention consisting of n-dodecyl pyrrolidone ("n-DP") in combination with alkyl polyglucoside ("APG") was added to solutions prepared from each of three commercially available enzyme cleaning preparation concentrates.

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These were MEDIZYME[®], ENDOZYME[®] and EPIZYME[®]. A fourth concentrate "NOVO formulation", a product described by NOVO INDUSTRIES in Novo Nordisk Publications Application Sheet B-613, "Application of Enzymes in Detergents for Endoscope Cleaning", was also prepared and tested. In the tests described in Table 1

5 the additive consisted of 0.2%w/w n-dodecyl pyrrolidone ("n-DP") and 0.1%w/w of alkyl polyglucoside ("APG") in each case based on the weight of the concentrate.

The NOVO Formulation (Concentrate) is as follows:

Component	%w/w
10 Non-ionic surfactant (Teric BL9)	12
Propylene Glycol	22
Triethanolamine	7
Boric acid	4
Calcium Chloride	0.1
15 Savinase 16.0L (Protease)	13
Termamyl (Amylase)	7
Water	to 100
pH	7

20 The Epizyme composition (concentrate) is as follows:

Component	%w/w
Tap water	19.0
Teric BL9 (non ionic surfactant)	10.0
Preservative	0.15
25 Enzyme stabilizer	6.0
50% caustic soda	2.5
Propylene Glycol	6.0
Protease	10.0
Lipase	0.2
30 Amylase	1.5
Cellulase	0.5
Perfume	0.15
Water	to 100.0

By way of example the protease may be subtilisin, the amylase Alpha-amylase, the cellulase may be endo-1,4-beta-glucanase and the lipase may be Triacylglycerol. However, other enzymes or combinations of enzymes may be substituted for these.

- 5 Each of the cleaning composition concentrates were diluted at a rate of 1 part of concentrate in 100 parts of standard hard water (0.304 g calcium chloride and 0.065g of magnesium chloride per 1L of distilled water. Therapeutic Goods Order 54 & Guidelines, 1996, p. 17). The efficacy of each product was then tested against standard soil preparations on glass slides, (1) neat and (2) with the addition of an additive
10 according to the invention. The efficiency of the preparation (time required to achieve a given score) was measured at various temperatures. The results are shown in Table 1 in comparison with results of compositions including the additive according to the invention.

- Accelerated aging tests are summarized in Table 2. Details of the preparation of
15 standard soils is given in table 3 and "Methods for Ascertaining the Cleaning Performance of Dishwasher Detergents" (SOFW-Journal, 126. Jahrgang 3-2000)

TABLE 1

Soil carrier glass microscope slides

- 20 Burnt minced meat. Contact time (min) required to achieve score 8.

	NOVO-formulation		EPIZYME		MEDIZYME		ENDOZYME	
	Neat	With APG + n-DP	neat	With APG + n-DP	neat	With APG + n-DP	neat	With APG + n-DP
25 °C	95	68	67	47	120	93	>480	241
40 °C	60	46	52	37	87	71	212	157
50 °C	42	33	30	19	65	53	170	140

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Milk in microwave. Contact time (in min) required to achieve score 10 (full removal)

	NOVO-formulation		EPIZYME		MEDIZYME		ENDOZYME	
	Neat	With APG + n-DP	neat	With APG + n-DP	neat	With APG + n-DP	neat	With APG + n-DP
25 °C	53	38	35	25	39	30	69	51
40 °C	39	31	24	19	29	23	50	35
50 °C	27	22	12	10	19	16	31	28

Edinburgh Soil. Contact time (in min) required to achieve score 10 (full removal)

	NOVO-formulation		EPIZYME		MEDIZYME		ENDOZYME	
	neat	With APG + n-DP	neat	With APG + n-DP	neat	With APG + n-DP	neat	With APG + n-DP
25 °C	28	25	21	19	26	23	32	28
40 °C	21	20	11	10	17	11.5	25	35
50 °C	18	15	9	9	11	10	18	16

- 5 N-DP: N-dodecyl pyrrolidone (Surfadone LP 300 from International Speciality Products),

APG: Alkyl polyglucoside (ALKADET 15 from Huntsman Corporation Australia)

Addition rate: 0.2 w/w% N-DP and 0.1 w/w% APG to concentrate

Dilution rate: 1 part of concentrate in 100 parts of standard hard water

10

As is evident from Table 1, the addition of the additive reduces the time required for the composition to achieve a given score and does so at each temperature. The additive is especially effective at lower temperatures.

- 15 Although the results shown in Table 1 were obtained using 0.2 w/w% N-DP and 0.1w/w% APG to concentrate, the ratio of N-DP to APG can be varied. It is preferred that the ratio be between 1 part by weight of alkyl pyrrolidone to 3 parts by weight of alkyl polyglucoside and 3 parts by weight of alkyl pyrrolidone to 1 part by weight of alkyl polyglucoside, more preferably between 3 parts by weight of alkyl pyrrolidone to 1
- 20 parts by weight of alkyl polyglucoside and 1 part by weight of alkyl pyrrolidone to 1 part by weight of alkyl polyglucoside.

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The concentration of N-DP and APG is each preferably less than 7.5% w/w of the concentrate (ie less than about .0075% in the concentrate at dilution of 1 part concentrate:100 parts water) and more preferably is less than 3% w/w. The actual concentrations employed will depend on the surface tension properties required, the hydrophilicity of the alkyl polyglucosides and the degree of hydrotrope provided by other excipients in the formulation to the alkyl polyglucoside, as well as other factors which may occur with respect to individual formulations.

Table 2 shows the effect of the additive in accelerated aging tests. The results demonstrate that the additive slightly improves storage stability, or, at least, does not adversely affect it.

TABLE 2

Remaining Protease Activity in % for concentrated formulations after 21 days storage (100% at day 0):

	NOVO-formulation		EPIZYME		MEDIZYME		ENDOZYME	
	neat	With APG + n-DP	neat	With APG + n-DP	neat	With APG + n-DP	neat	With APG + n-DP
25 °C	95	95	96	98	95	95	93	93
40 °C	89	89	91	91	85	85	80	83
47 °C	77	84	85	87	69	74	73	78

Assessed using B 863b test method by NOVO NORDISK (attached herewith as Annexure 1) The additive according to the invention and dilution were as for table 1.

Table 3 shows the effects of varying the concentration and ratio of the alkylpyrrolidone and alkylpolyglucoside. It can be seen that the composition is efficacious in reducing contact times for cleaning over a wide range of alkylpyrrolidone to alkylpolyglucoside ratios.

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TABLE 3

Soil carrier: glass microscope slides

Burnt minced meat. Contact time (min) required to achieve score 8.

	NOVO-formulation		Epizyme	
	neat	With APG + SURF	Neat	With APG + SURF
25 °C	95	66	67	48
40 °C	60	49	52	39
50 °C	42	32	30	22

5 Milk in microwave. Contact time (in min) required to achieve score 10 (full removal)

	NOVO-formulation		Epizyme	
	neat	With APG + SURF	Neat	With APG + SURF
25 °C	53	34	35	21
40 °C	39	29	24	17
50 °C	27	20	12	8

Edinburgh Soil. Contact time (in min) required to achieve score 10 (full removal)

	NOVO-formulation		Epizyme	
	neat	With APG + SURF	neat	With APG + SURF
25 °C	28	22	21	19
40 °C	21	16	11	9
50 °C	18	19	9	8

SURF: n-dodecyl pyrrolidone (Surfadone LP 300 from International Speciality

10 Products),

APG: Alkyl polyglucoside (ALKADET 15 from Huntsman Corporation Australia)

Addition rate: 0.2 w/w% SURF and 1% APG to concentrate

Dilution rate: 1 part of concentrate in 100 parts of standard hard water

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Results.

Soil carrier: glass microscope slides

Burnt minced meat. Contact time (min) required to achieve score 8.

	NOVO-formulation		Epizyme	
	neat	With APG + SURF	neat	With APG + SURF
25 °C	95	64	67	44
40 °C	60	47	52	37
50 °C	42	31	30	22

5 Milk in microwave. Contact time (in min) required to achieve score 10 (full removal)

	NOVO-formulation		Epizyme	
	neat	With APG + SURF	neat	With APG + SURF
25 °C	53	34	35	21
40 °C	39	28	24	16
50 °C	27	20	12	7

Edinburgh Soil. Contact time (in min) required to achieve score 10 (full removal)

	NOVO-formulation		Epizyme	
	neat	With APG + SURF	neat	With APG + SURF
25 °C	28	21	21	19
40 °C	21	15	11	9
50 °C	18	14	9	7

SURF: n-dodecyl pyrrolidone (Surfadone LP 300 from International Speciality

10 Products),

APG: Alkyl polyglucoside (ALKADET 15 from Huntsman Corporation Australia)

Addition rate: 0.2 w/w% SURF and 7.5% APG to concentrate

Dilution rate: 1 part of concentrate in 100 parts of standard hard water

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Results.

Soil carrier: glass microscope slides

Burnt minced meat. Contact time (min) required to achieve score 8.

	NOVO-formulation		Epizyme	
	neat	With APG + SURF	neat	With APG + SURF
25 °C	95	67	67	44
40 °C	60	44	52	41
50 °C	42	30	30	20

5 Milk in microwave. Contact time (in min) required to achieve score 10 (full removal)

	NOVO-formulation		Epizyme	
	neat	With APG + SURF	neat	With APG + SURF
25 °C	53	37	35	24
40 °C	39	26	24	15
50 °C	27	18	12	12

Edinburgh Soil. Contact time (in min) required to achieve score 10 (full removal)

	NOVO-formulation		Epizyme	
	neat	With APG + SURF	neat	With APG + SURF
25 °C	28	18	21	19
40 °C	21	15	11	12
50 °C	18	12	9	7

SURF: n-dodecyl pyrrolidone (Surfadone LP 300 from International Speciality

10 Products),

APG: Alkyl polyglucoside (ALKADET 15 from Huntsman Corporation Australia)

Addition rate: 1% w/w% SURF and 0.1% APG to concentrate

Dilution rate: 1 part of concentrate in 100 parts of standard hard water

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Results.**Soil carrier: glass microscope slides****Burnt minced meat. Contact time (min) required to achieve score 8.**

	NOVO-formulation		Epizyme	
	neat	With APG + SURF	neat	With APG + SURF
25 °C	95	63	67	49
40 °C	60	52	52	37
50 °C	42	35	30	23

5 Milk in microwave. Contact time (in min) required to achieve score 10 (full removal)

	NOVO-formulation		Epizyme	
	neat	With APG + SURF	neat	With APG + SURF
25 °C	53	35	35	24
40 °C	39	27	24	16
50 °C	27	24	12	9

Edinburgh Soil. Contact time (in min) required to achieve score 10 (full removal)

	NOVO-formulation		Epizyme	
	neat	With APG + SURF	neat	With APG + SURF
25 °C	28	23	21	20
40 °C	21	17	11	9
50 °C	18	12	9	7

SURF: n-dodecyl pyrrolidone (Surfadone LP 300 from International Speciality**10 Products),****APG: Alkyl polyglucoside (ALKADET 15 from Huntsman Corporation Australia)****Addition rate: 7.5% w/w% SURF and 0.1% APG to concentrate****Dilution rate: 1 part of concentrate in 100 parts of standard hard water****Assessed using B 863B test method by NOVO NORDISK (attached)**

TABLE 4

SOIL COMPOSITION & DETERGENT ENZYME ACTIVITY TEST**Milk in Microwave:** - 60 ml of milk

- 6 beakers

5 - a microwave set to 450 W

- an oven heater set to 80 C°

The microwave was pre-heated by placing 6 water-filled beakers in a circle on the glass turn-table and leaving it on "high" (900 W) for 10 minutes. Then, the beakers with water were replaced by beakers with 10 ml of milk. The milk was heated for 10 minutes at 450 W, then allowed to cool, forming a skin, and placed into an oven for 2 hours at 80°C.

Mince meat: -20 grams of beef mince

-7.5 grams of egg yolk and whites

- 8.0 gram of water

15 30x30 mm polar substrate (ceramic tiles)

The mince was mixed with the egg yolk and water with the use of a hand blender. 1g of homogenized mixture was spooned onto 3x3 cm ceramic tiles, onto the smooth side. Then the tiles were left in an oven for 10 minutes.

Edinburgh Soil - 20 g egg yolk

20 - 20 g water

- 0.5 g cholesterol

- 3 g beef gelatin

- 0.45 hog mucin

The beef gelatin and water were blended together and heated until the mixture obtained a homogenous consistency. Cholesterol, hog mucin and egg yolk were blended in after gelatin was cooled to 30C. 0.1 g of soil was deposited onto surfaces of the microscope slides. The smears were made of a uniform thickness for each set of slides; the thickness was regulated by a border of adhesive tape along each border of the smearing instrument. In order to simulate the conditions of an endoscope cleaning, the slides were left to dry in open air at 20°C for 20 minutes.

Without wishing to be bound by theory, the polysaccharide appears to stabilize the pyrrolidone, the combination being more soluble in the concentrate and resulting in substantially improved lowering of surface tension at the working dilution.

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Although the test results utilize n-dodecyl pyrrolidone, other alkyl pyrrolidones could be substituted therefore, and especially C8 to C18 alkyl pyrrolidones. The amount to be added can be determined by simple trial but in general increasing the concentration significantly above the levels herein employed tends to give a decreasing advantage.

- 5 Similarly, other alkyl polysaccharides may be substituted for APG. For example other sugar units and derivatives of sugar units may be substituted for glucose units. Again there is an optimum concentration beyond which increasing the concentration is of relatively little benefit. While the combination of n-alkyl pyrrolidone with APG is highly preferred, beneficial effects are obtainable with at least some soils by adding it in
10 isolation. The optimum ratio of n-DP to APG can be determined by simple trial for a given concentrate.

- The combination of the invention provides an improved surfactant system which is expected to have application in many end-uses. In the field of cleaners for medical instruments, the surfactant system of the invention may be simply added to
15 commercially available enzyme cleaning products either to the concentrate or, at an appropriate dilution to the working solution prior to use. Preferably however the additives are incorporated into a concentrate at the time of manufacture of the concentrate which is then merely diluted prior to use.

- Additives according to the invention may be added to dishwashing, hand
20 washing and other surface cleaning compositions.

- Embodiments of the invention may be altered and may have other chemicals added or substituted therefore to an extent which will be apparent to those skilled in the art from the teaching hereof without departing from the inventive concept herein
25 disclosed.

ANNEXURE 1.**NOVO NORDISK B 863b-GB****5 MANUAL PROCEDURE FOR DETERMINATION OF PROTEOLYTIC ACTIVITY IN ENZYME PREPARATIONS AND DETERGENTS (AZOCASEIN SUBSTRATE)**

A diagrammatic representation of this method is shown in figure 1.

Principle

- 10 A protease is allowed to hydrolyse azocasein for 30 minutes at 40°C. Undigested protein is precipitated with trichloroacetic acid and the quantity of digested product is determined by spectrophotometry.

Enzyme Unit Definition

- 15 No separate unit is defined. Activity is determined relative to a standard of known activity and the result given in the same units as used for the standard.

Assay Conditions

- pH: 8.5 (0.2 M Tris-SO₄ buffer)
Temperature: 40°C
Substrate: 0.5% azocasein (in reaction mixture)
20 Reaction time: 30 minutes

Apparatus

- Spectrophotometer to read absorbance at 390 nm
pH meter
Water bath at 40°C ± 0.5°C
25 Timer with a second hand
Vortex mixer
16 • 125 mm test tubes
Repeating dispenser, 5 ml capacity
Small funnels and filter paper, Whatman no. 3 or equivalent

- 15 -

Reagents**1. 2.0 M Tris buffer stock solution**

Dissolve 242 g trishydroxymethylaminomethane (Sigma "Trizma Base" T-1503 or equivalent) in 700-800 ml deionised water.

5 Adjust pH to 8.5 with 10 N H₂SO₄.

Adjust volume to 1 litre.

This concentrate is to be used at 10 ml/100 ml volume for all final dilutions with 0.2 M Tris buffer.

10 2. 50% urea solution

Dissolve 50 g urea (analytical grade) in 50 ml warm deionised water.

Adjust volume to 1 litre.

3. 10% trichloroacetic acid solution (stop reagent)**15 Dissolve 100 g trichloroacetic acid (TCA) in 200 ml deionised water.**

Adjust volume to 1 litre.

4. Azocasein (substrate solution)

Make fresh daily - discard any unused substrate

20 In a 250-ml beaker:

Weigh 0.6g azocasein (Sigma A-2765).

Add 10 ml 50% urea solution and mix until dissolved.

Add 10 ml 2.0 M Tris buffer stock solution.

Add 30-50 ml deionised water and continue stirring until clear of particles.

25 Adjust pH to 8.5 using dilute H₂SO₄. Adjust volume to 100 ml and mix thoroughly.

Keep cool until ready to begin assay.

5. Enzyme samples and standards

See Appendix I for Alcalase®, Appendix II for Esperase® and Savinase®, and Appendix III for Durazym®.

30

After dissolving a detergent sample for protease assay, it is strongly recommended that the pH of this solution be checked and, if needed, adjusted to 8.5 • 0.1 units.

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If it is desired to know the exact activity levels of proteinase in a detergent, the detergent bases should be included in the standard solutions unless it is already known that the particular formulation being used has no effect on the protease activity. The concentration of the base detergent should be equal to that used in the sample solution of the detergent containing protease.

The presence in solution of strong chelators such as NTA or EDTA may result in deactivation of the protease over the time period required for preparation of the various materials for the assay. This cause of deactivation can be overcome by adding excess calcium to the sample solutions.

Procedure

1. Preheat water bath to $40^{\circ}\text{C} \pm 0.5^{\circ}\text{C}$ (approx. 1 hr.).
2. Approx. 10 minutes before starting assay, place substrate solution in 40°C water bath to equilibrate.
3. To appropriately labelled test tubes, pipet 1 ml of sample or enzyme standard and equilibrate to 40°C in water bath (approx. 1 minute).
4. Start reaction:
At precisely timed intervals, add 5 ml substrate solution to each tube containing sample or standard, vortex and return to water bath for 30 minutes.
5. Stop reaction:
After exactly 30 minutes, at the same time intervals as before, add 5 ml of 10% TCA to each sample or standard tube, vortex and let stand at room temperature
6. After 15-20 minutes filter all tubes by gravity filtration through paper filters into clean, dry and properly -labelled test tubes.
7. Read absorbances at 390 nm vs deionised water blank.

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Calculations

1. Standard curve:

a) If standard blank (A_0 , E_0 or S_0) > 0.16 , assay should be re-run with fresh substrate

5 b) If standard blank < 0.16 , subtract its A_{390} readings from standards to obtain absorbance

c) Plot a curve of • absorbance vs proteolytic activity (AU or KNPU/tube = AU or KNPU/ml of final dilution).

10 2. Sample tubes:

$$\bullet \text{ absorbance} = A_{\text{sample}} - A_{\text{blank}}$$

Using standard curve, determine activity of sample/tube,

$$\text{Activity/gram} = \frac{(\text{activity/tube}) \bullet (\text{dilution factor})}{\text{gram of sample}}$$

15

APPENDIX 1

Alcalase®

5 Unit definition: Anson Unit (AU)

1 Anson Unit = the amount of enzyme* which digests haemoglobin A at an initial rate such that there is liberated per minute an amount of TCA-soluble product which gives the same colour with phenol reagent as one milliequivalent of tyrosine.

10 * Under reaction conditions given in the NNAS method AF4/5-GB: Modified Anson-Haemoglobin Method for the Determination of Proteolytic Activity.

A standard curve is prepared using an enzyme whose activity in Anson Units is known. Unknowns are then compared to this curve.

15

Buffer: 0.2 M Tris, pH 8.5

Sample activity range: $5 \cdot 10^{-5}$ -- $15 \cdot 10^{-5}$ AU/ml

Standard blank: 1 ml of 0.2 M Tris buffer pH 8.5 = A_0

Substrate: Azocasein solution, 0.5%, pH 8.5

20 Sample preparation:

Dissolve an accurately-weighed amount of sample in an accurately-known volume of 0.2 M Tris buffer so that the solution contains between $5 \cdot 10^{-5}$ and $15 \cdot 10^{-5}$ AU/ml and has a pH of 8.5.

25

N.B. For each unknown, prepare a sample blank and one or more duplicates (as necessary).

Keep solutions on ice until ready to assay.

Standard curve

30 Enzyme standard stock solution: Dissolve accurately-weighed Alcalase standard in 100 ml 0.2 M Tris buffer (pH 8.5) to yield $2.6 \cdot 10^{-3}$ AU/ml

Dilute the stock solution as follows:

35

Tube 1 label	Stock solution	2 M Tris buffer stock	Final volume	Activity
A_0	0 ml	10 ml	100 ml	$0.0 \cdot 10^{-5}$ AU/ml
A_1	1 ml	10 ml	100 ml	$2.6 \cdot 10^{-5}$ AU/ml
40 A_2	2 ml	10 ml	100 ml	$5.2 \cdot 10^{-5}$ AU/ml
A_3	3 ml	10 ml	100 ml	$7.8 \cdot 10^{-5}$ AU/ml
A_4	4 ml	10 ml	100 ml	$10.4 \cdot 10^{-5}$ AU/ml
A_5	5 ml	10 ml	100 ml	$13.0 \cdot 10^{-5}$ AU/ml
45 A_6	6 ml	10 ml	100 ml	$15.6 \cdot 10^{-5}$ AU/ml

APPENDIX II

Savinase®/Esperase®

5 Unit definition: Kilo Novo Protease Unit (KNPU)

1 KNPU = 1000 NPU

1 Novo Protease Unit (NPU) = the amount of enzyme* which hydrolyzes casein at such a rate that the initial rate of formation of peptides/minute corresponds to 1 micromole of glycine/minute

10

* Standard conditions given in NNAS method 162/3-GB: Manual DMC Method for the Determination of Proteolytic Activity.

A standard curve is prepared using an enzyme whose activity in KNPU) is known.

15 Unknowns are then compared to this curve

Buffer: 0.2 M Tris, pH 8.5

Sample activity range: $2 \cdot 10^{-4}$ – $6 \cdot 10^{-4}$ KNPU/ml

Standard blank: 1 ml of 0.2 M Tris buffer pH 8.5 = E_0 or S_0

20 Substrate: Azocasein solution, 0.5%, pH 8.5

Sample preparation:

Dissolve an accurately-weighed amount of sample in an accurately-known volume of 0.2 M Tris buffer so that the solution contains between $2 \cdot 10^{-4}$ – $6 \cdot 10^{-4}$ KNPU/ml and

25 has a pH of 8.5.

N. B. For each unknown, prepare a sample blank and one or more duplicates (as necessary).

Keep solutions on ice until ready to assay.

30 Standard curve

Enzyme standard stock solution: Dissolve accurately-weighed Savinase or Esperase standard in 100 ml 0.2 M Tris buffer (pH 8.5) to yield $1 \cdot 10^{-2}$ KNPU/ml.

Dilute the stock solution as follows:

35

Tube	Stock solution	2 M Tris buffer stock	Find volume	Activity
E_0 or S_0	0 ml	10 ml	100 ml	$0.0 \cdot 10^{-4}$ KNPU/ml
40 E_1 or S_1	1 ml	10 ml	100 ml	$1.0 \cdot 10^{-4}$ KNPU/ml
E_2 or S_2	2 ml	10 ml	100 ml	$2.0 \cdot 10^{-4}$ KNPU/ml
E_3 or S_3	3 ml	10 ml	100 ml	$3.0 \cdot 10^{-4}$ KNPU/ml
E_4 or S_4	4 ml	10 ml	100 ml	$4.0 \cdot 10^{-4}$ KNPU/ml
E_5 or S_5	5 ml	10 ml	100 ml	$5.0 \cdot 10^{-4}$ KNPU/ml
45 E_6 or S_6	6 ml	10 ml	100 ml	$6.0 \cdot 10^{-4}$ KNPU/ml

E = Esperase, S = Savinase

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APPENDIX III

Durazym®

Unit definition: Durazym Protease Unit (DPU)

- 5 The activity is measured relative to a Durazym standard with the same units as the sample i.e. DPU.

A standard curve is prepared using an enzyme whose activity in DPU is known. Unknowns we then compared to this curve

- 10 Buffer: 0.2 M Tris, pH 8.5

Sample activity range: $6 \cdot 10^{-4}$ – $18 \cdot 10^{-4}$ DPU/ml

Standard blank: 1 ml of 0.2 M Tris buffer pH 8.5 = D_0

Substrate: Azocasein solution, 0.5%, pH 8.5

Sample preparation:

- 15 Dissolve an accurately-weighed amount of sample in an accurately-known volume of 0.2 M Tris buffer so that the solution contains between $6 \cdot 10^{-4}$ – $18 \cdot 10^{-4}$ DPU/ml and has a pH of 8.5.

- 20 N.B. For each unknown, prepare a sample blank and one or more duplicates (as necessary).

Keep solutions on ice until ready to assay.

Standard curve

Enzyme standard stock solution: Dissolve accurately-weighed Durazym standard in

- 25 100ml 0.2 M Tris buffer (pH 8.5) to yield $1 \cdot 10^{-2}$ DPU/ml

Dilute the stock solution as follows:

30	Tube label	Stock solution	2 M Tris buffer stock	Final volume	Activity
	D_0	0 ml	10 ml	100 ml	$0.0 \cdot 10^{-4}$ DPU/ml
	D_1	3 ml	10 ml	100 ml	$3.0 \cdot 10^{-4}$ DPU/ml
	D_2	6 ml	10 ml	100 ml	$6.0 \cdot 10^{-4}$ DPU/ml
35	D_3	9 ml	10 ml	100 ml	$9.0 \cdot 10^{-4}$ DPU/ml
	D_4	12 ml	10 ml	100 ml	$12.0 \cdot 10^{-4}$ DPU/ml
	D_5	15 ml	10 ml	100 ml	$15.0 \cdot 10^{-4}$ DPU/ml
	D_6	18 ml	10 ml	100 ml	$18.0 \cdot 10^{-4}$ DPU/ml

40

THE CLAIMS OF THE INVENTION ARE AS FOLLOWS:

1. An aqueous composition comprising in combination an alkyl pyrrolidone and an alkyl polysaccharide.
5
2. A composition according to claim 1 wherein the alkyl pyrrolidone is a C8 -C18 linear alkyl pyrrolidone.
3. A composition according to claim 1 or 2 wherein the alkyl polysaccharide is an
10 alkyl polyglucoside ("APG").
4. A composition according to any one of the preceding claims wherein the alkyl pyrrolidone is n-dodecyl pyrrolidone ("n-DP").
- 15 5. A composition according to any one of the preceding claims wherein the ratio of alkyl pyrrolidone to alkyl polysaccharide is between a ratio of 1 part by weight of alkyl pyrrolidone to 3 parts by weight of alkyl polyglucoside and 3 parts by weight of alkyl pyrrolidone to 1 part by weight of alkyl polyglucoside.
- 20 6. A composition according to any one of the preceding claims wherein the ratio of alkyl pyrrolidone to alkyl polysaccharide is between a ratio of 1 part by weight of alkyl pyrrolidone to 3 parts by weight of alkyl polyglucoside and 1 parts by weight of alkyl pyrrolidone to 1 part by weight of alkyl polyglucoside.
- 25 7. A composition according to any one of the preceding claims comprising about 1 part by weight of n-dodecyl pyrrolidone to about 1 part by weight of alkyl polyglucoside.
8. A method of enhancing the efficacy of an enzyme containing composition for use
30 in cleaning medical instruments comprising the step of including in said composition an alkyl pyrrolidone and an alkyl polysaccharide.

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9. A method according to claim 8 wherein a combination according to any one of claims 1 to 6 is added to an enzyme containing composition.
10. A method according to claim 9 wherein an alkyl pyrrolidone and an alkyl polysaccharide are separately added to an enzyme containing composition.
11. A method according to any one of claims 9 to 10 wherein the enzyme containing composition is a concentrate and the alkyl pyrrolidone and an alkyl polysaccharide are added to the concentrate prior to its dilution to a working strength.
12. A method according to claim 8 wherein an alkyl pyrrolidone and an alkyl polysaccharide are each formulated in combination with one or more enzymes to produce a concentrate suitable for dilution to provide an enzyme cleaning solution.
13. A method of cleaning a medical instrument including the step of treating the instrument with a composition including at least one enzyme, an alkyl pyrrolidone and an alkyl polysaccharide.
14. A method according to claim 13 wherein the alkyl pyrrolidone is a C8 -C18 linear alkyl pyrrolidone.
15. A method according to claim 13 or 14 wherein the alkyl polysaccharide is an alkyl polyglucoside ("APG").
16. A method according to any one of claims 13 to 15 wherein the alkyl pyrrolidone is n-dodecyl pyrrolidone ("n-DP").
17. A method according to any one of claims 13 to 16 wherein the ratio of alkyl pyrrolidone to alkyl polysaccharide is between a ratio of 1 part by weight of alkyl pyrrolidone to 3 parts by weight of alkyl polyglucoside and 3 parts by weight of alkyl pyrrolidone to 1 part by weight of alkyl polyglucoside.
18. A method according to any one of claims 13 to 17 wherein the ratio of alkyl pyrrolidone to alkyl polysaccharide is between a ratio of 1 part by weight of alkyl

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pyrrolidone to 3 parts by weight of alkyl polyglucoside and 1 parts by weight of alkyl pyrrolidone to 1 part by weight of alkyl polyglucoside

19. A method according to any one of claims 12 to 15 comprising about 1 part by weight of n-dodecyl pyrrolidone to about 1 part by weight of alkyl polyglucoside.

20. A composition according to any one of claims 1 to 7 further including at least one enzyme.

21. A composition according to claim 20 including at least one enzyme selected from the group consisting of proteases, lipases, amylases, and cellulases.

22. A composition according to claim 21 including at least two different enzymes each selected from the group consisting of proteases, lipases, amylases, and cellulases.

23. A composition according to any one of claims 20 to 22 further comprising propylene glycol.

24. A composition according to any one of claims 20 to 23 further comprising a non ionic surfactant.

25. A composition according to any one of claims 20 to 24 wherein APG is present in a concentration of less than 7.5% w/w.

26. A composition according to any one of claims 20 to 25 wherein n-DP is present in a concentration of less than 7.5% w/w.

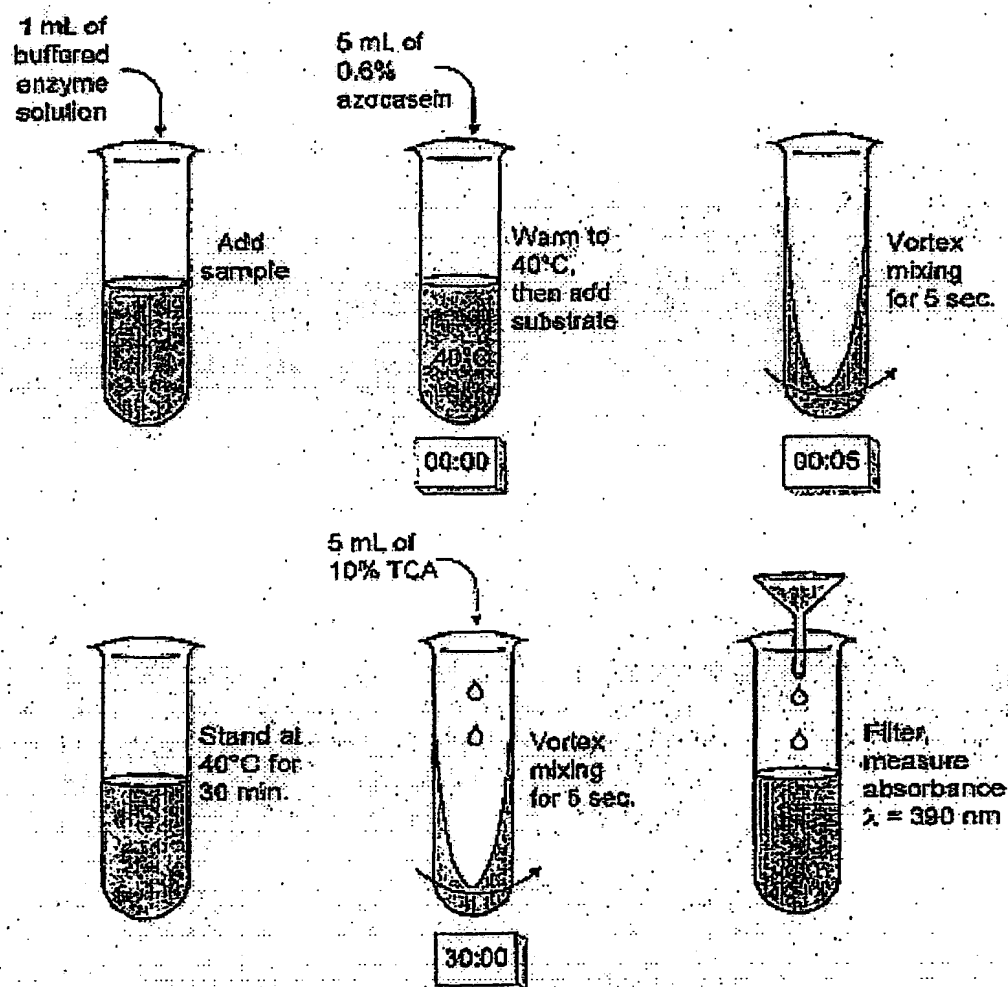
25. A composition according to any one of claims 20 to 24 wherein APG is present in a concentration of less than 0.3% w/w.

26. A composition according to any one of claims 20 to 25 wherein n-DP is present in a concentration of less than 0.3% w/w.

27 A composition substantially as herein described with reference to any one of the examples

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FIG. 1



INTERNATIONAL SEARCH REPORT

International application No.

PCT/AU01/01094

A. CLASSIFICATION OF SUBJECT MATTER		
Int. Cl. 7: C11D 7/32, A61L 2/18		
According to International Patent Classification (IPC) or to both national classification and IPC		
B. FIELDS SEARCHED		
Minimum documentation searched (classification system followed by classification symbols) SEE "ELECTRONIC DATABASE" BOX BELOW		
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched SEE "ELECTRONIC DATABASE" BOX BELOW		
Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) WPIDS, JAPIO, CA: ?pyrrolidone, ?polyglucoside, ?polysaccharide, detergent, clean?		
C. DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 6 040 288 (POPOFF et al.) 21 March 2000 column 10, paragraph 3	1
X	US 5 922 663 (GABRIEL et al.) 13 July 1999 column 6, paragraph 3	1
X	US 5 908 619 (SCHOLZ) 1 June 1999 column 7, paragraph 2; column 19, paragraph 3	1
<input checked="" type="checkbox"/> Further documents are listed in the continuation of Box C <input checked="" type="checkbox"/> See patent family annex		
<p>* Special categories of cited documents:</p> <p>"A" document defining the general state of the art which is not considered to be of particular relevance</p> <p>"B" earlier application or patent but published on or after the international filing date</p> <p>"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</p> <p>"O" document referring to an oral disclosure, use, exhibition or other means</p> <p>"P" document published prior to the international filing date but later than the priority date claimed</p> <p>"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</p> <p>"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone.</p> <p>"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art</p> <p>"&" document member of the same patent family</p>		
Date of the actual completion of the international search 19 September 2001		Date of mailing of the international search report 27 SEPTEMBER 2001
Name and mailing address of the ISA/AU AUSTRALIAN PATENT OFFICE PO BOX 200, WODEN ACT 2606, AUSTRALIA E-mail address: pct@ipaaustralia.gov.au Facsimile No. (02) 6285 3929		Authorized officer JAMIE TURNER Telephone No : (02) 6283 2071

INTERNATIONAL SEARCH REPORT

International application No.

PCT/AU01/01094

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 5 863 886 (TRACY et al.) 26 January 1999 claim 12	1
X	US 5 846 926 (TRACY et al.) 8 December 1998 claim 17	1
X	US 5 558 806 (POLICELLO et al.) 24 September 1996 claim 4; example 2	1-2

INTERNATIONAL SEARCH REPORT
Information on patent family members

International application No.
PCT/AU01/01094

This Annex lists the known "A" publication level patent family members relating to the patent documents cited in the above-mentioned international search report. The Australian Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

Patent Document Cited in Search Report			Patent Family Member				
US	6060288	CA	2196604	EP	784701	US	5641658
		US	6090592	WO	9604404	AU	61460/98
		WO	9836094				
US	5922663	AU	43931/97	EP	951346	WO	9815346
US	5908619	AU	53858/98	EP	963157	WO	9830096
US	5863886	NONE					
US	5846926	NONE					
US	5558806	NONE					
END OF ANNEX							

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